#### Course Unit Descriptor

Study Programme: Doctoral Academic Studies in Biochemistry

Course Unit Title: Cell cultures in medicinal chemistry

Course Unit Code: DSB-624

Name of Lecturer(s): Professor Suzana Jovanović-Šanta

Type and Level of Studies: PhD degree

Course Status (compulsory/elective): elective

Semester (winter/summer): summer

Language of instruction: English

Mode of course unit delivery (face-to-face/distance learning): Face-to-face

Number of ECTS Allocated: 15

**Prerequisites:** -

# Course Aims:

The aim of the course is to acquire integrated knowledge about the possibilities of applying cell model-systems, as well as the ability of scientifically interpreted experimental data in the field of in vitro testing of the biological activity of potential therapeutics.

## Learning Outcomes:

After successfully completing the course, students should be able to use the methods and techniques necessary to perform in vitro tests on cell cultures as model systems, and have the ability to critically analyze scientific papers, scientific hypotheses, and experimental results in in vitro assays on cell cultures, and independently know how to perform an experiment related to the given area and process and discuss the results obtained.

## Syllabus:

Theory

Maintenance of cell lines. Biology of cell cultures, primary and continuous cell lines. Culture of tumor and control cells (healthy tissue cells). Physical methods for cell separation. Quantification and characterization of cell growth. *In vitro* tests with cell cultures: measurement of cell viability, antiproliferative and cytotoxic activity; methods for detecting and quantifying cell death and cell cycle phases; studying changes in cell morphology by specific colors: changes on the membrane, DNA, organelles, identification of proteins characteristic of individual signal pathways by modern molecular methods; use of cell cultures to assess the genotoxic effects of compounds or extracts.

### Practice

Preclinical studies of the pharmacological potential of compounds by qualitative and quantitative *in vitro* methods: Measurement of cell viability, morphological changes using specific color dyes, cell cycle changes, apoptosis measurement, necrosis and membrane potential of mitochondrial by flow cytometry, genotoxicity testing (SCE, MN and other tests). Study of available data and development of a mini-project to investigate the pharmacological potential of substances or mixtures in cell cultures as a model system.

### **Required Reading:**

- 1. Freshney R. I. (ed) (2005) Culture of animal cells: a manual of basic technique. 5th ed.Wiley-Liss. New York, USA.
- 2. D.A. Gewirtz, S.E. Holt, S. Grant (2007) Apoptosis, Senescence, and Cancer, Humana Press Inc (Eds)
- 3. C. Avendano, J.C. Menendez (2008) Medicinal chemistry of anticancer drugs, Elsevier
- 4. Review and original scientific articles from selected research areas

Weekly Contact Hours: 10		Lectures: 5		Practical work: 5
Teaching Methods: Lectures, laboratory work, discussion of original research articles, study projects				
Knowledge Assessment (maximum of 100 points): 100				
Pre-exam obligations	points		Final exam	points
Active class			written exam	
participation			witten exam	
Project presentation	50		oral exam	50
Preliminary exam(s)				
Seminar(s)				
The methods of knowledge assessment may differ; the table presents only some of the options: written exam, oral exam,				
project presentation, seminars, etc.				